THE EFFECT OF ADMINISTRATION OF ETHANOL EXTRACT FROM MUSA PARADISIACA L. (MPL) FRUIT ON THE CASPASE-3 MRNA GENE EXPRESSION IN RAT AMYLOID BETA INDUCED, AN ALZHEIMER'S DISEASE MODEL

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THE EFFECT OF ADMINISTRATION OF ETHANOL EXTRACT FROM *MUSA PARADISIACA* L. (MPL) FRUIT ON THE CASPASE-3 MRNA GENE EXPRESSION IN RAT AMYLOID BETA INDUCED, AN ALZHEIMER'S DISEASE MODEL

EMMA KAMELIA[1,3], ANDI ASADUL ISLAM[2], MOCHAMMAD HATTA[3]*, CAHYONO KAELAN[4], ILHAMJAYA PATELLONGI[5]

[1]Dental Health Study Program, Tasikmalaya Health Polytechnic, West Java, Indonesia. [2]Department of Neurosurgery, Medical Faculty, University of Hasanuddin, Makassar, Indonesia. [3]Molecular Biology and Immunology Laboratory for Infection Diseases, Medical Faculty, University of Hasanuddin, Makassar, Indonesia. [4]Department of Neurology, Medical Faculty, University of Hasanuddin Makassar, Indonesia. [5]Department of Physiology, Medical Faculty, University of Hasanuddin, Makassar, Indonesia. Email: hattaram@yahoo.com

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ABSTRACT

Objective: The objective of this study is to analyze caspase-3 mRNA gene expression in a Wistar rat model of Alzheimer's disease (AD) treated with *Musa paradisiaca* L. (MPL) ethanol extract or banana extract (BE)

Methods: MPL was evaluated for its effect on the caspase-3 mRNA gene expression of rat amyloid beta (Aβ) induced, an AD model. Each model included twenty Wistar rats were randomized into five groups: K0, without AD induction and no BE; K1, AD induction and no BE; K2, AD induction+BE 250 mg/kg body weight (BW); K3, AD induction+BE 500 mg/kg; and K4, AD induction+BE 1000 mg/kg. AD induction was performed by $Aβ_{1-42}$ (0.2 ug) injection at the intracerebroventricular area. mRNA caspase-3 level measurements were performed by real time-polymerase chain reaction.

Results: Paired t-test analysis showed no significant differences of caspase-3 mRNA level before Aβ induction among five groups (p>0.05). At 6 weeks post-Aβ induction, there was significantly increased caspase-3 mRNA in all groups except K0 (p<0.05). Notably, after 3 weeks of BE administration, caspase-3 mRNA expression was significantly decreased in all BE-treated groups; in the K1 placebo group, caspase-3 mRNA expression increased. The maximum decreased caspase-3 mRNA expression was in group K4 (-BE 1000 mg/kg BW), and the minimum was in group K2 (-BE 250 mg/kg BW).

Conclusion: The results revealed that the ethanolic extract of MPL fruit could decrease caspase-3 mRNA gene expression in AD rat

Keywords: Alzheimer's disease, Intracerebroventricularly, Amyloid beta, Caspase-3, *Musa paradisiaca* L.

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INTRODUCTION

Alzheimer's disease (AD) is an irreversible and progressive neurodegenerative disease, a form of dementia known globally, projected that, by 2050, about 115 million people will be affected worldwide [1]. As the aging population increases, the number of patients with AD also increases, with a rate of 13% in older patients over 65 years old and 45% in the group aged over 85 years old [2]. The majority of AD cases occur in women compared with men, and the estimated risk for developing AD is about 20% for women and 10% for men age above 65 [3]. AD was initially discovered by German neural expert Alois Alzheimer in 1906, with found brain lesions that lead to dementia and they were highly associated with strokes, brain tumor, and degenerative diseases [4]. Several factors contribute to the progression of the disease including amyloid-beta (Aβ) accumulation, neurofibrillary tangle formation, cholinergic deficit, oxidative stress, neuroinflammation, and apoptosis [5].

Apoptotic signaling is classified as proceeding by either an intrinsic pathway or an extrinsic pathway. The intrinsic apoptotic signaling is most often induced by the intracellular damage that leads to the mitochondrial release of cytochrome c and the activation of intracellular cysteine proteases called caspases [6]. Extrinsic apoptotic signaling is initiated by stimulation of plasma membrane death receptors that initiate apoptosis by activation of caspase-8, and subsequent apoptotic signaling can proceed through the mitochondrial pathway or independently of mitochondria by caspase-8-mediated direct activation of caspase-3 [7]. Caspase-3 acts as a caspase executor in the apoptosis process [8]. Caspase-3 appears to be the major effector in neuronal apoptosis triggered by various stimuli. The first strong evidence supporting the specific role for this protease in neuronal apoptosis comes from studies on mice deficient in caspase-3 in which brain development is profoundly altered [9]. Furthermore, Aβ secretion decreased when there is an obstacle to its production in caspase-3 caused by antioxidant and pramipexole [10]. The drugs designed to slow disease progression are available. Some medicinal herbs from plants may help to improve brain function, but scientific evidence to prove that they can treat AD is limited.

Commenges *et al.* found that an intake of flavonoids significantly reduced the risk of dementia [11]. In this research, caspase-3 mRNA gene expression in an AD rat model induced by injection of Aβ for 6 weeks, which was treated with banana extract (BE)/*Musa paradisiaca* L. (MPL) for 3 weeks, was evaluated. The purpose of this study was to analyze the effect of BE (MPL) administration on the expression of the caspase-3 mRNA gene in the Wistar rats model of AD, with the hypothesis that BE MPL administration for 3 weeks can decrease the expression of the caspase-3 mRNA gene in Wistar rats which is in Aβ induction for 6 weeks. BE referred to herein is BE or MPL which is extracted using ethanol.

METHODS

Materials

Experimental procedures were carried out in the Molecular Microbiology and Immunology Laboratory, Medical Faculty University

of Hasanuddin, Makassar, Indonesia. This research was an experimental study *in vivo* pre and post-design that was conducted from February to July 2016. The research has been approved by the Medical and Health Research Ethics Committee, Medical Faculty University of Hasanuddin, Makassar, Indonesia (Number: 391/H4.8.4.5.31/PP36-KOMTIK/2016).

Animals

Animal preparation

Twenty Wistar rats (2–3 months, 150–250 g) were maintenance at Animal Laboratory, Molecular Biology and Immunology Laboratory, Medical Faculty University of Hasanuddin, Makassar, Indonesia, for this study. The rats were randomly assigned to one of five groups (=4 in each group): The control (K0 and K1) and treatment groups (K2, K3, and K4). They were kept for 1 week for proper acclimatization before starting the experiment under the controlled condition of illumination (12 h light/12 h darkness) and temperature 23±2°C. They were housed under ideal laboratory conditions and maintained on the standard pellet diet and water *ad libitum* throughout the experimental period. All procedures were by the internationally accepted guideline of experimental animal use and care of Animals Laboratory of the Molecular Microbiology and Immunology Laboratory, Medical Faculty University of Hasanuddin, Makassar.

Experimental design

In this study, total number of twenty Wistar rats were divided into following five groups having four rats in each group: K0=no induction AD model and no BE MPL administration; K1=AD induction model without BE MPL administration (just given a placebo); K2=AD induction model with 250 mg/kg body weight (BW) BE MPL administration; K3=AD induction model with 500 mg/kg BW BE MPL administration; and K4=AD induction model with 1000 mg/kg BW BE MPL administration. Caspase-3 mRNA expression was assessed before AD induction, 6 weeks after AD induction, and 3 weeks post-BE MPL administration.

Administration of MPL ethanol extract BE

Banana fruits were obtained from a market in Bogor, West Java Province, Indonesia. The fruits were cut longitudinally into chips of about 5–10 mm thickness and then dried in an oven after which they were ground and added ethanol (95%) to a conical flask and then shaken every 30 min (for 6 h) and sterilized for 48 h. The solution is then filtered using filter paper, ethanol added to the solution coming out of the colorless outer macerator (usually 5–6 times immersion) and then concentrated using a rotary evaporator until no more solvent, which drips on the rotary evaporator condenser. This is called solid dry extract and is used in this study. Three doses of the plantain flour were prepared: 250 mg/kg/day, 500 mg/kg/day, and 1000 mg/kg/day. The flour was dissolved in 2 ml of double-distilled water, for easy administration. Banana MPL ethanol extract or BE was obtained by maceration methods at Balitro Bogor Agricultural Department Laboratory, West Java Province, Indonesia, and plant determination process from Central Biology Research, Bogor LIPI, West Java Province, Indonesia (Number: 147/IPH.1.02/lf.07/I/2016). Previous research by Ittiyavirah and Anurenj showed that a 200 mg/kg BW dose of MPL ethanol extract BE given over 21 days produced a significant effect on anti-stress activity [12]. In the study for AD, we increased the dose to 250 mg/kg, and the BE used on K2 is 250 mg/kg BW, K3 500 mg/kg BW, and K4 1000 mg/kg BW for experimental analyses; treatments were given orally for 21 days (3 weeks) each day. While in K1 only given a placebo with the same time of administration is 3 weeks. Overall, both BE MPL and placebo were administered by mouth-feeding.

Animal model

In this research, we do advance preliminary research which was done to develop an animal model of the AD. To develop an AD model, we used a dose of $A\beta_{1\text{-}42}$ of 0.2 µg injected in intracerebroventricular and observed the mice for 6 weeks, and Aβ levels were measured using the SAA Mouse Elisa Kit. The AD animal model was generated using $A\beta_{1\text{-}42}$ peptide from Abcam, code number.Ab120959 (Cambridge, MA, USA) [13], which is a reference modeling AD protocol.

Blood collection

Blood samples were collected at three time points: Day 0 before Aβ injection (baseline); day 42 (week: 6), i.e., on the last day of post-AB induction observation or the day before the MPL intervention; and last is the day 63, the last day of BE intervention which done in 21 days from day 42 (weeks: 9, changes). Blood was taken from the tail vein using a 0.1 ml hematocrit needle. The samples were centrifuged and were kept in a sterile tube at −20°C until analyses.

Real time-polymerase chain reaction (RT-PCR)

The RT-PCR Method uses the Boom method, which has high sensitivity and specificity for disease diagnosis. In the extraction of DNA, the materials used are diatoms, L6 (lysis buffer), L2 (washing buffer), and TE (elution buffer). The diatoms have large contours that will bind DNA. While L6 serves as a lysis buffer and L2 as a washing buffer, and Tris EDTA (TE) to bind DNA out of diatoms. The RT-PCR was carried out to determine the levels of caspase-3 mRNA gene expression. RT-PCR was performed with the following primers, caspase-3 forward: 5'-AGCTTCTTCAGAGGCGACTA-3' and reverse: 5'-GGACACAATACACGGGATCT-3' [15]; and glycerol-aldehyde-3-phosphate dehydrogenase (GAPDH) forward: 5'-CTCAAGATTGTCAGCAATGC-3' and reverse: 5'- CAGGATGCCCTTTAGTGGGC-3' [16]; reactions were performed using the one-step RT-PCR kit (Macrogen, Korea), GADPH was used as an internal control. Primers for RT-PCR were designed using Oligo software (Bio-Rad CFX Manager). The blood samples from treatment and control in homogenization and total RNA samples in extraction according to the instructions of the protocol. Reactions were visualized by agarose gel electrophoresis analysis. Rate threshold cycle in the calculation of the standard software. The level of expression exraction according to the instructions of the protocol. Making PCR mix: using solution (Table 1), for caspase-3 made in triplicate. Then take 5.2 µl caspase-3 into each well (each well requires a 5.2 µl PCR mix). Also, distribute GAPDH (as standard) into the wells, each well needs a 5.2 µl PCR mix. Here, fill the samples in the wells each containing the PCR mix, caspase-3, and GAPDH, wait for 1.5 h. Then, turn on the PCR tool and make a layout, then input the sample that has been idle for 1.5 h into the wells, each enter as much as 5.2 µl, neither with the standard (GAPDH). Next is dilution: 20 µ RNA free water inserted into the new tube (tubes 1, 2, 3, 4, 5, and 6), then the DNA template of all samples takes as much as 20 µ and diluted with 20 µ which had been diluted and so on up to the six tubes. Then, enter 5 µ to the wells that have been made earlier, close and insert into the PCR machine, perform an initial denaturation stage of 96°C for 3 min, next 95°C for 30 s, next annealing 55°C for 30 s with volume contents 5+5=10 µ 40 times.

Statistical analysis

The data caspase-3 mRNA gene expression was statistically analyzed and the significance calculated using one-way "ANOVA" (Bonferroni) followed by Tukey's test. All numerical values were expressed as a mean±standard deviation and the value of $p<0.05$ was considered as statistically significant.

Fig. 1 Schematic of the research process. Twenty Wistar rats were divided into 5 groups: K0=no induction AD model and no BE administration; K1=AD induction model without BE administration; K2=AD induction model with 250 mg/kg BW BE administration; K3=AD induction model with 500 mg/kg BW BE administration; and K4=AD induction model with 1000 mg/kg BW BE administration. Caspase-3 mRNA expression

Table 1: Stages of PCR mix manufacturing

RT-PCR	Caspase-3 (×80)	GAPDH (×30)
Master mix=2.5 µl	200 µl	75 µl
Free water=2.5 µl	200 µl	75 µl
Primer Caspase-3 F=0.1 µl	8 µl	3 µl
Primer Caspase-3 R=0.1 µl	8 µl	3 µl
Template=5 µl		

RT-PCR: Real time-polymerase chain reaction

was assessed before AD induction, 6 weeks after AD induction, and 3 weeks post-BE administration.

RESULTS

In this study, we examined the effects of MPL administration at various doses in an AD animal model. The schematic of our research strategy is shown in Fig. 1. The levels of caspase-3 mRNA at various time points throughout the analysis were evaluated. The caspase-3 mRNA expression levels in all observation groups are summarized in Table 2

Paired t-test and one-way ANOVA (Bonferroni) showed changes in caspase-3 mRNA expression after 6 weeks post-induction Aβ, and no significant differences in caspase-3 mRNA expression among all five groups before induction were observed (p>0.05). At 6 weeks post-induction Aβ, there was a significant increase (p<0.05) in caspase-3 mRNA expression in all groups except the K0 group. There were no significant differences in the increase in caspase-3 mRNA expression among induction groups (p>0.05). At 6 weeks after induction Aβ, we detected increased caspase-3 mRNA expression in the induction Aβ groups, while no changes occurred in the non-induction Aβ group (K0).

Paired t-test and one-way ANOVA (Bonferroni) were used to examine changes in caspase-3 mRNA expression at 3 weeks post-BE admission, and after 3 weeks of BE administration, there was a significant increase in caspase-3 mRNA in the group without BE (placebo) compared with the levels at week before BE treatment (p<0.05). Notably, in all groups receiving BE, all showed a significant decrease in caspase-3 mRNA expression compared with the levels at week 6 before BE treatment (p<0.05). After 3 weeks of BE admission during the post-induction period, caspase-3 mRNA expression decreased in all groups given BE. In comparison, in the group given the only placebo, the caspase-3 mRNA levels continued to increase. The maximum decrease in caspase-3 mRNA expression was observed in the K4 group, which received a BE dosage of 1000 mg/kg BW. The smallest decrease was observed in group K2, which received a BE dosage of 250 mg/kg BW.

DISCUSSION

Despite extensive investigations, the mechanism activating apoptotic pathways or neuronal degeneration in the AD has not been completely understood. In the present study, we showed the effects of administration of MPL extract BE on caspase-3 mRNA expression in Aβ-induced AD rats. Our results show that: (i) in both K1 (placebo) and K2, K3 and K4 before MPL- detected treatment intervention experienced an increase in caspase-3 mRNA expression at 6 weeks after induction of Aβ, which is similar to the result presented by cetin *et al*. Their research found high caspase-3 activity levels in hippocampus, temporal, and parietal cortex in aged mice injected with $A\beta_{1-42}$ [17]. Aβ has been proposed as the main factor in the AD pathophysiology mechanism and the major

Table 2: Caspase-3 mRNA expression in each group

Variable	Groups	Observation time				
		Before induction (baseline week 0)	6 weeks after induction (weeks 6)	3 weeks post-BE treatment (Week 9)	0-6 p*	6-9 p
Caspase-3 mRNA expression mean±SD	K0 (n=4)	7.68 (0.25)^{A}	7.66 (0.23)^{A}	7.64 (0.06)^{A}	0.900	0.871
	K1 (n=4)	7.65 (0.07)^{A}	12.69 (0.16)^{B}	13.67 (0.16)^{B}	<0.001	0.007
	K2 (n=4)	7.54 (0.38)^{A}	12.49 (0.07)^{B}	11.54 (0.05)^{D}	<0.001	<0.001
	K3 (n=4)	7.63 (0.07)^{A}	12.73 (0.15)^{B}	9.23 (0.18)^{C}	<0.001	<0.001
	K4 (n=4)	7.68 (0.10)^{A}	12.53 (0.04)^{B}	8.65 (0.10)^{C}	<0.001	<0.001

Values are mean±SD. Statistics+one-way ANOVA followed by Bonferroni *post hoc* test, *p>0.05 versus K0, *p<0.05 versus K1, *p<0.05 versus K2, *p<0.05 versus K3, *p<0.05 versus K4. N=Number of rats in each group, BE: Banana extract, SDM=Standard deviation mean

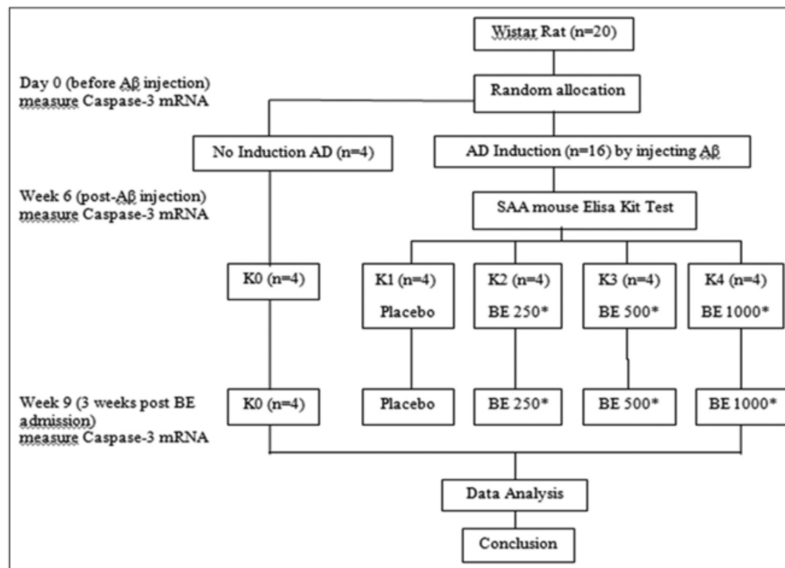

Fig. 1: Outline of research process

component of senile plaques, is considered to play a central role in neuronal cell death (apoptosis), and has received the most attention [18]. This has been demonstrated in neurons of human and rodent brains. In patients suffering from neurodegenerative disorders such as AD, PD, and amyotrophic lateral sclerosis, there is a prominent increase in the neuronal RNA damage when compared to normal aging people [19]. Several factors that are assumed to contribute in AD pathogenesis, such as the role of key proteins, oxidative damage, mitochondrial dysfunction, and the cholinergic hypothesis [20]. The role of neuronal cell death (apoptosis) includes caspase-9 and caspase-3 [21]. Our next findings in this study are (ii) that the placebo group continued to show significantly increased caspase-3 mRNA levels for the next 3 weeks. However, in groups treated with MPL BE, we observed the opposite trend, with a significant decrease of caspase-3 mRNA expression ($p<0.05$). The maximum decreased level of caspase-3 mRNA expression was observed in K4, the group which received the highest dose of MPL BE in this study (1000 mg/kg BW BE), and the minimum decrease was observed in group K2, which received 250 mg/kg BW BE. This result is consistent with the study by Vijayakumar *et al.*, who reported findings of flavonoid antioxidant MPL activity in mice [22]. The antioxidant activity of the extracted flavonoids from *M. paradisiacal* in rats stimulated the activities of superoxide dismutase and catalase which might be responsible for the reduced level of peroxidation products such as malondialdehyde, hydroperoxides, and conjugated dienes [22].

The authors stated that flavonoids could modulate the expression of several genes through activation of many transcription factors and act as a neuroprotection agent, were thought to work by modulating kinase proteins, and act as a kinase cascade lipid signal, such as phosphatidyl inositol-3 kinase (PI3K)/Akt pathway signal, tyrosine kinase, protein kinase C (PKC) and mitogen-activated protein kinase (MAPK), and PKB [23]. Activation of neuronal cell membrane receptors may result in caspase activation, enhanced calcium levels, and the generation of reactive oxygen species (ROS), both are known to contribute to the mitochondrial pathway, including B-cell lymphoma 2 (Bcl-2) proteins, high intracellular calcium levels. and ROS. On entering the cytoplasm, cytochrome c forms an apoptosomal complex with a procaspase, adenosine triphosphate, and apoptotic protease activating factor 1 (Apaf-1), an Apaf-1 [21]. The apoptotic pathway is generally characterized by various signals including mitochondrial release from proapototic molecules such as Bcl-2, ROS, high calcium and cytochrome C binds Apaf-1 and procaspase 9 to form apoptosomes, which further activate caspase 9nd procaspase 9 to form the apoptosome, thereby activating caspase 9. Caspase-9 activates caspase, ultimately leading to cell death [24]. Heo *et al.* used BE MPL to protect neurons from oxidative stress by neurotoxicity. The study also showed that BE MPL could reduce oxidative stress risk caused by neurodegenerative diseases, such as AD [22]. The BE possess very good radical scavenging activity and as well the largest amount of phenolic contents, which could introduce phenols as the main radical scavenger in BE and offering effective protection from free radicals and the antioxidant activity [25]. The difference with the previous research that they only see MPL on oxidative stress and as an antioxidant, but in this study, emphasized its effect on the expression of the caspase-3 mRNA gene. We add the biochemical evidence showing that treatment with ethanolic extract MPL can decrease serum expression of caspase-3 mRNA in an animal model for the AD, suggesting that it may prevent cell apoptosis. Further studies are required to isolate and evaluate individual phytoconstituents in MPL for their neuroprotection potential.

CONCLUSION

Taking together, it may finally be concluded that Aβ-induced AD causes neuron cell death, where apoptosis plays an important role in it and may be inhibited by flavonoids, may exert regulatory activities in cells through actions such as cyclin-dependent kinases, caspases, Bcl-2 family members, epidermal growth factor/epidermal growth factor receptor, phosphatidylinositol-3-kinase/Akt, MAPK, and include the inhibition of c-Jun N-terminal kinases and p38 pathways and the activation of PI3-K/Akt, and PKC pathways in different type of cell such as neuronal, cardiac, endothelial, epithelial, hepatocytes, and macrophages. Such interactions of flavonoids with cell signaling pathways provide various beneficial effects such as improving brain function, preventing oxidative stress, preventing apoptosis, protecting against endothelial barrier dysfunction and injury, improving the cognitive function, decreasing the neurodegeneration, and stimulating endothelial NOS activity. That natural flavonoids can inhibit caspase-3 activity. Medication using MPL ethanol extract can decrease caspase-3 mRNA expression in AD rats. Suggesting that these flavonoids may be useful which leads to the rational design of non-peptidic, caspase-specific inhibitors for therapeutic use. Herbs may play a promising role in the early treatment of Alzheimer's and other conditions involving poor memory and dementia. One of the chief benefits is that they have a low side effect compared to pharmaceutical agents.

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CONTRIBUTION

The first author has carried out the research. Second, third, fourth, and fifth authors have provided study conception, the design of work, drafting of the manuscript, and critical revision.

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